Chromosome 19:
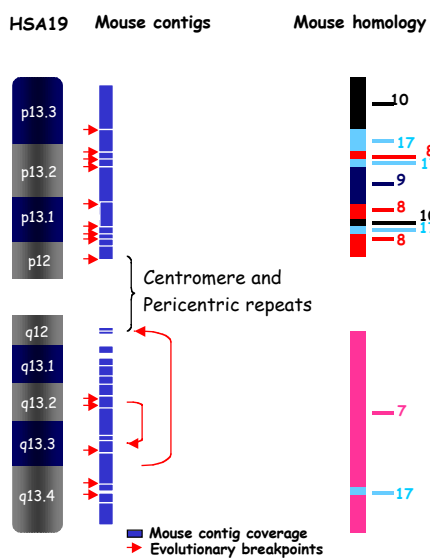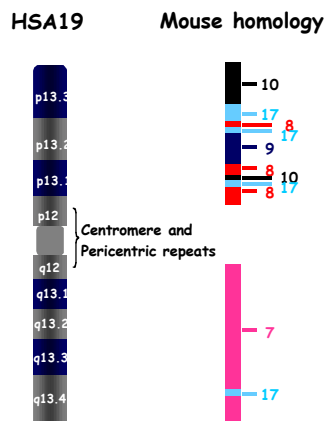
- ~65-70 Mb total length
    - estimate of up to 1100 genes
    - ~17 Mb centromere + pericentromeric repeats (few or no genes)
    - ~2 Mb gene "deserts"
    - 46 Mb gene containing regions targeted for comparative sequencing

- 57 Mb contiguous clone map with 7 gaps
    - 35 Mb finished sequence
    - 22 Mb mostly o&oed draft

- 15 homology segments related to Mmu7, 8, 9, 10 and 17

HSA19   Mouse homology

p13.3
p13.2
p13.1
p12
q12
q13.1
q13.2
q13.3
q13.4

Centromere and Pericentric repeats

10
17
8
17
9
8
10
8
17

7

17

HSA19   Mouse contigs   Mouse homology

p13.3
p13.2
p13.1
p12

Centromere and Pericentric repeats

q12
q13.1
q13.2
q13.3
q13.4

10
17
8
17
9
8
10
17
8

7

17

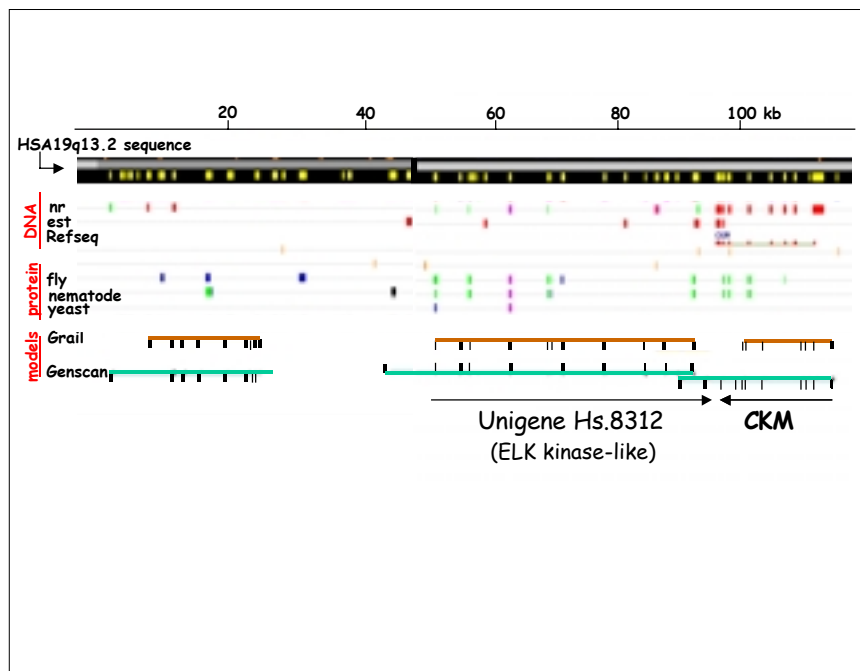■ Mouse contig coverage
➤ Evolutionary breakpoints

Status

~35 mouse BAC contigs spanning the lengths of all 15 homology segments

- several homology segments spanned by a single contig
- > 95% coverage of mouse chromosome 19-related regions
- breakpoints of all evolutionary rearrangements cloned

>42 Mb non-overlapping mouse draft sequence completed
- All clones sequenced at $\geq$ 6X depth in paired plasmid ends
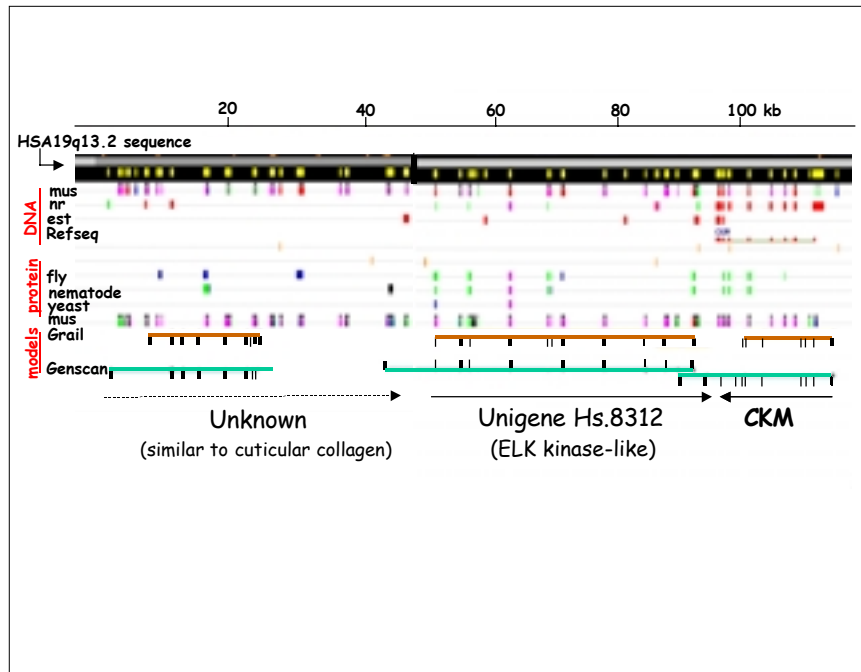- Sequence of >60% clones is fully ordered and oriented; most remaining are partially ordered

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Initial analyses focused on three major questions:

- Human sequence annotation:
    - value of comparative alignments for gene finding and functional-element definition
- Chromosome evolution:
    - what clues are provided by analysis of sequence at breaks in syntenic homology?
- Gene evolution:
    - How do primate and rodent gene sets compare?
        - What impact might species-specific differences have on biology?

Value of comparative sequence alignment as a sequence-annotation strategy
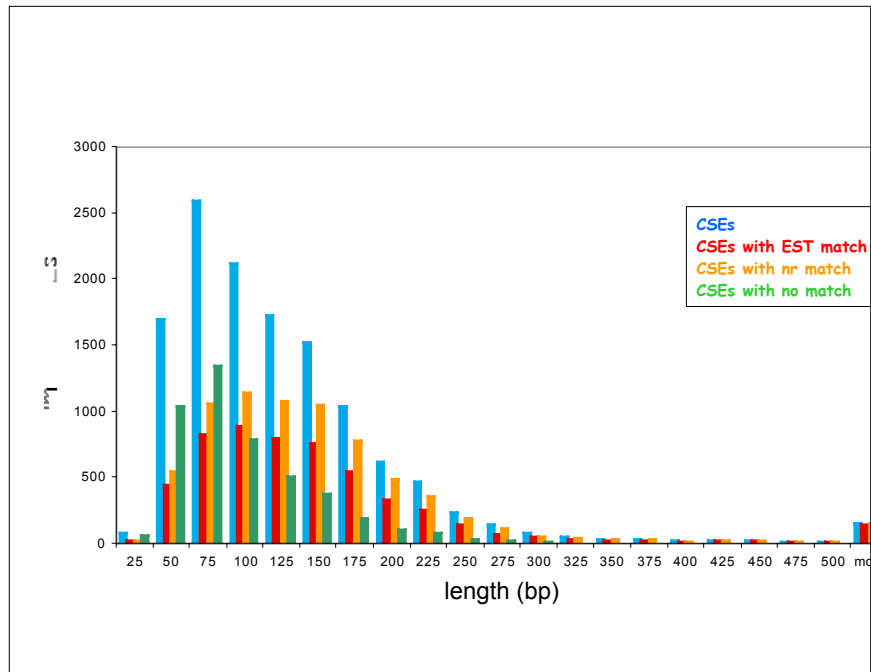
- What did we gain by sequencing mouse?
  - Identification of many new candidate exons -- 5' ends, alternative exons, etc. in known genes
  - Confirmation and expansion of predicted genes
  - Prediction of ~30 new candidate genes that would have been missed entirely by other gene-finding methods
  - 128 genes identified by EST + mouse conservation only
  - >4000 non-coding conserved sequences that are candidates for regulatory DNA sequence elements

Slide 1

20 40 60 80 100 kb

HSA19q13.2 sequence

DNA
mus
nr
est
Refseq

protein
fly
nematode
yeast
mus

models
Grail
Genscan

Unknown
(similar to cuticular collagen)

Unigene Hs.8312
(ELK kinase-like)

CKM

Slide 2

HSA19 conserved sequences

- **5.4%** of HSA19 sequence is conserved at significant levels in **syntenically homologous** mouse DNA
  - aligning non-homologous mouse sequence yields more conserved elements, but most are *not* functionally significant
- **80%** of the exons of known HSA19 genes are conserved in homologous regions of mouse
- **12611** conserved sequence elements:
  - **42%** coincide with high-probability EST matches
  - **57%** have significant match to non-redundant nucleotide or protein database entries
  - **36%** conserved sequences (4546 CSEs) do not coincide with any other sequence feature (EST, protein, nt, exon model)
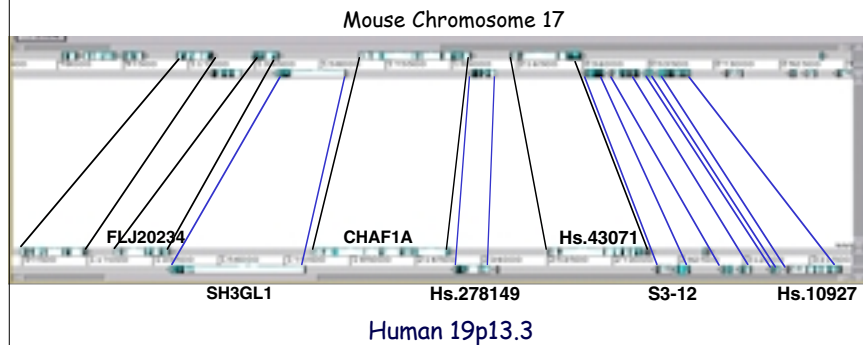
Gene number predictions and general observations

- Combining mouse sequence matches with other evidence we predict ~1200 HSA19 genes
  - ~860 (~70%) "unique" (or small-family) genes
  - ~340 (~30%) members of large clustered families
    - ZINC FINGER GENES, OLFACTORY RECEPTORS, VOMERONASAL RECEPTORS, CYTOCHROME P450 GENES, NATURAL KILLER RECEPTORS, SERINE PROTEASES, PREGNANCY SPECIFIC GLYCOPROTEINS, SIALIC ACID GYCOPROTEINS…...

- All but 30 predicted genes based on high probability EST matches

- Computer-based gene finding programs found one or more exons in ~55% of HSA19 genes (60% of known)

III. Conservation of human and mouse gene sets

Unique HSA19 genes are overwhelmingly conserved in mouse...

- Of 781 established (Refseq, Locus link, unigene) HSA19 genes, clear relatives were found for 744 (95%) in related mouse BAC sequence
  - 31 genes fall into gaps in the mouse BAC map (4.1%)
  - 3 genes are missing from well-covered mouse regions
    - PPPAR1A: member of gene distributed family
    - 2 unigene matches encoding hypothetical proteins

- In general, orthologous genes are arranged in identical order in homologous regions of HSA19 and mouse DNA…

Mouse Chromosome 17

FLJ20234   CHAF1A   Hs.43071

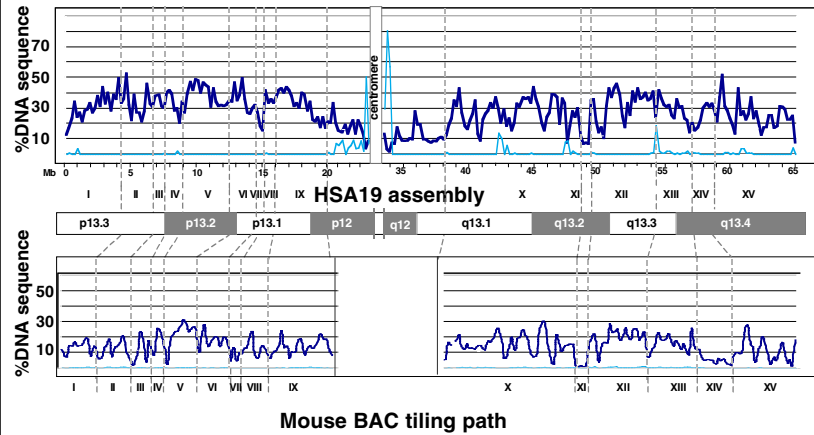SH3GL1   Hs.278149   S3-12   Hs.10927

Human 19p13.3

....But in regions totaling ~50% of HSA19 DNA, human genes are larger and more widely spaced due to a significant increase in the numbers of inserted SINE repeats

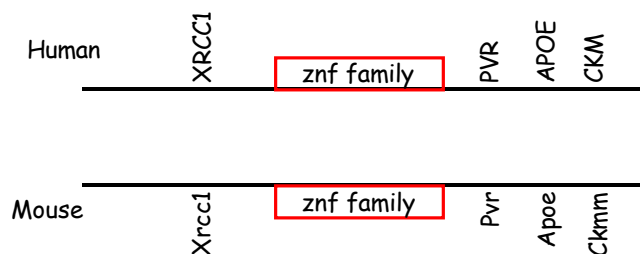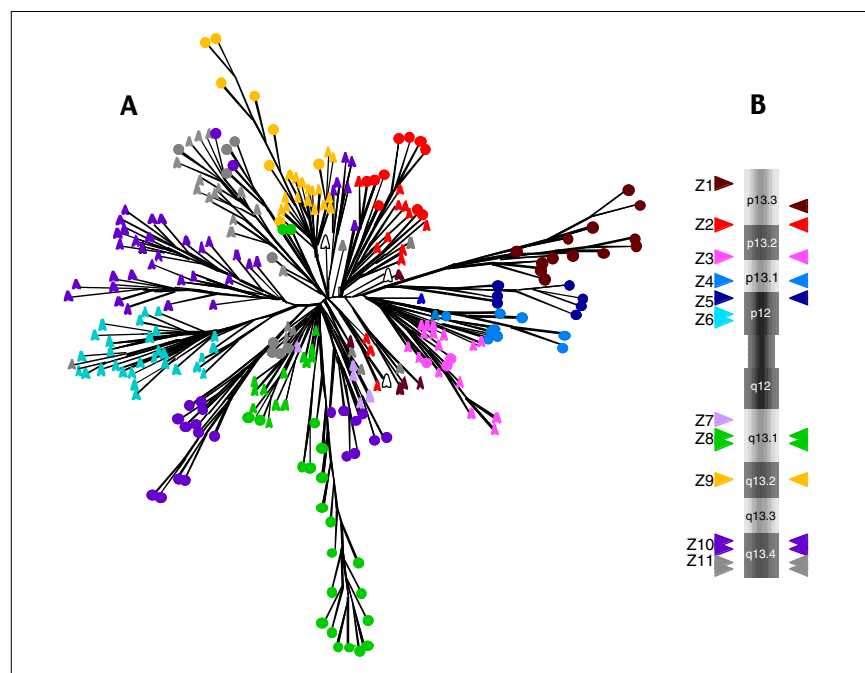Sines as percentage of total sequence, plotted along HSA19 and mouse
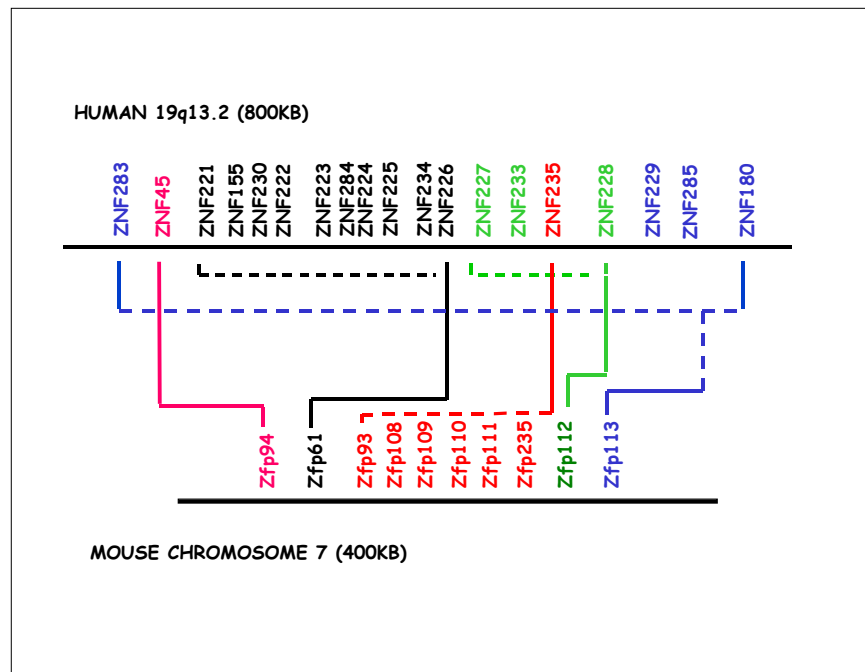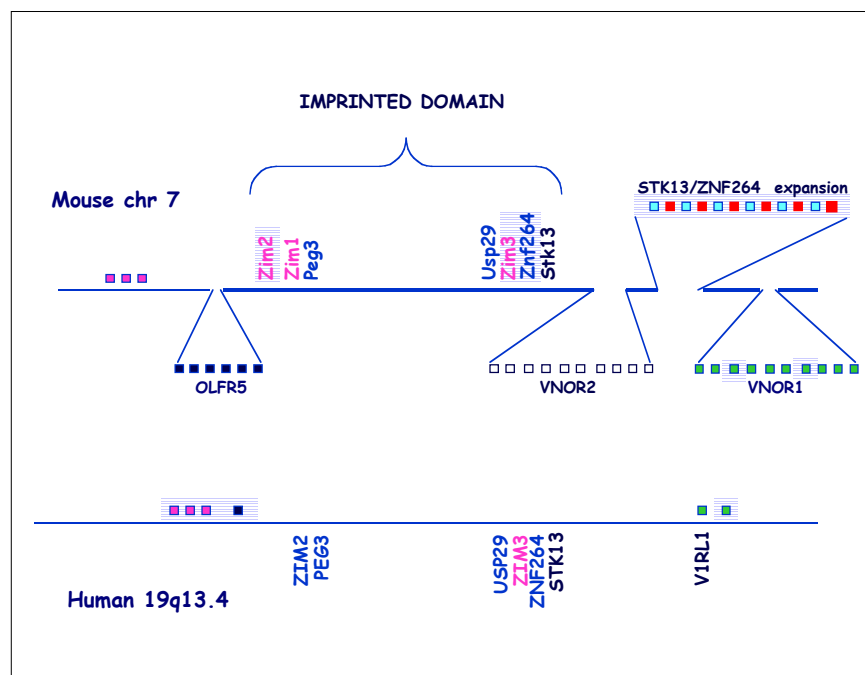
HSA19 average: ~27%   mouse: 12.7%
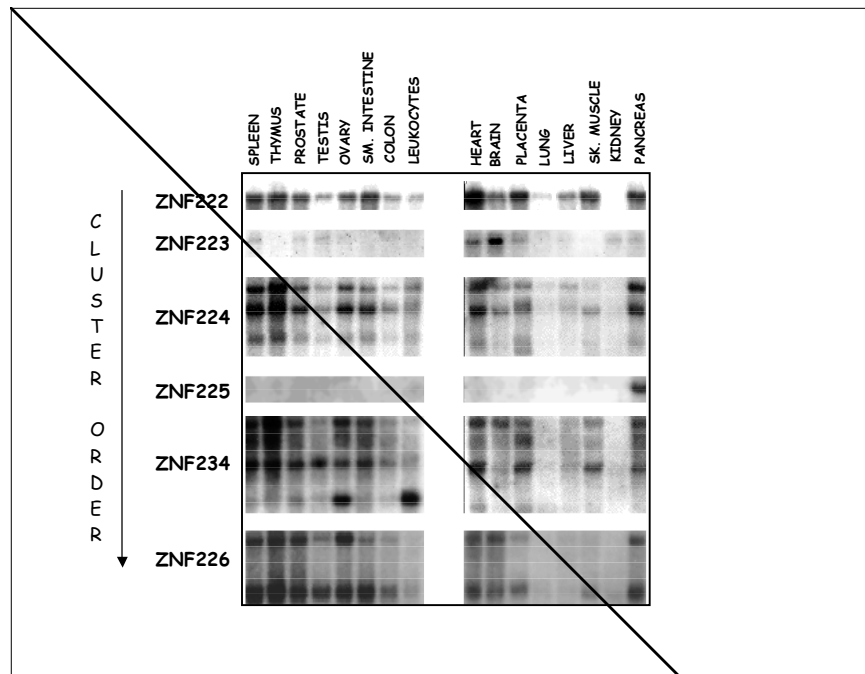

HSA19 assembly

Mouse BAC tiling path

...Tandem gene families

- MOST CLUSTERED HUMAN FAMILIES ARE REPRESENTED BY A RELATED FAMILY IN THE SYNTENICALLY CONSERVED POSITION

Human   XRCC1 | znf family | PVR APOE CKM

Mouse   Xrcc1 | znf family | Pvr Apoe Ckmm

HUMAN 19q13.2 (800KB)
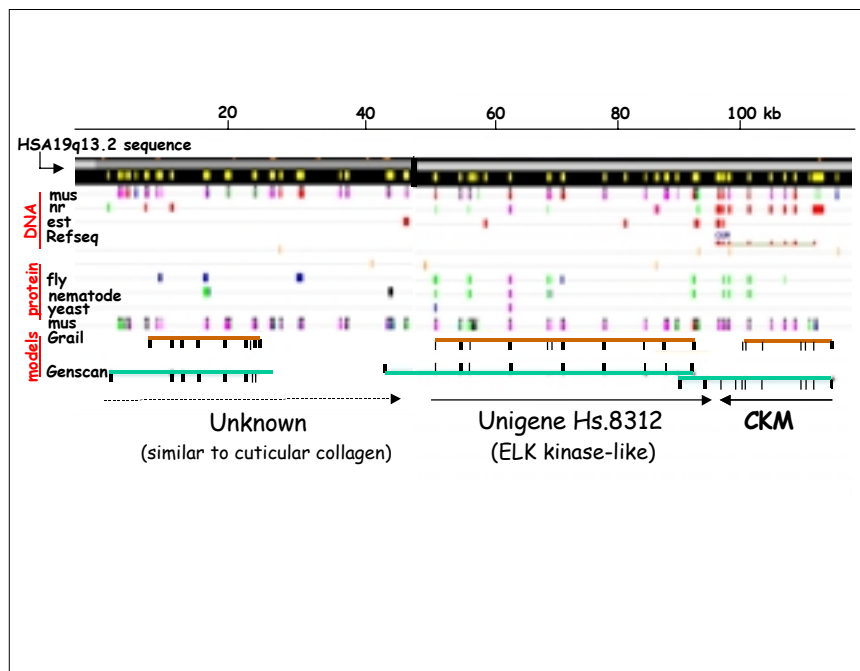
MOUSE CHROMOSOME 7 (400KB)

A

B

In general...

- In contrast to unique genes regions, tandemly clustered families differ extensively in gene content, gene number and organization between the two species
  - VNO receptors, OLFR genes: multiple functional copies in mouse, and multiple pseudogenes in ch19; human singletons represented by large clusters in mouse
  - ZNF genes: conserved clusters, but different gene complements due to ongoing differential gain and loss of gene copies
  - Many actively expressed, and probably functional, lineage-specific genes exist within these and other families, at least 100 on HSA19 alone
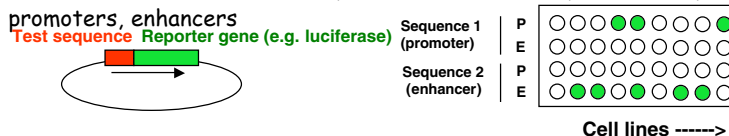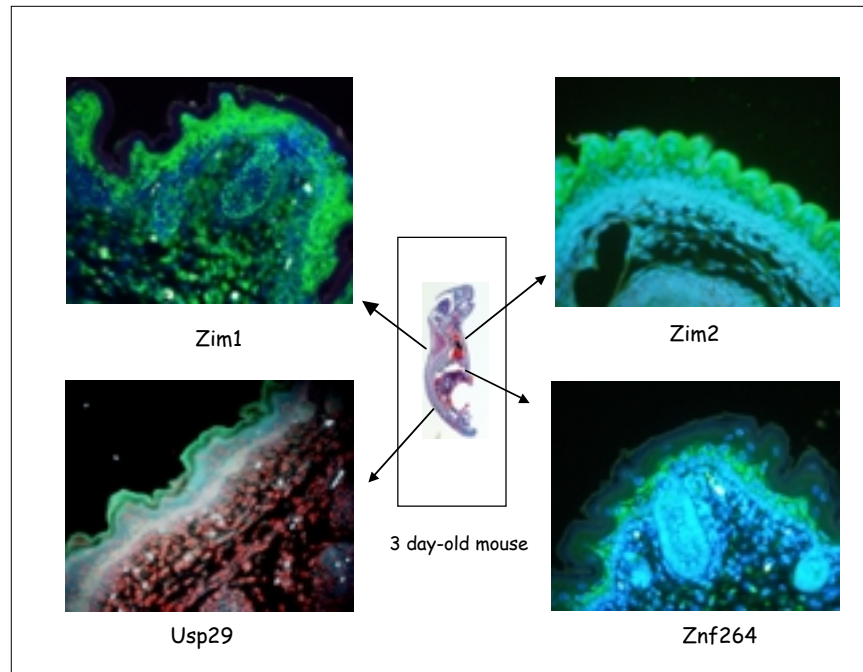

Unknown
(similar to cuticular collagen)

Unigene Hs.8312
(ELK kinase-like)

CKM

What's next?

- Defining the borders of known and predicted genes
  - Which elements are linked together to create specific transcription units? Are alternative transcripts generated in different tissues?
- Identifying and testing regulatory elements predicted by comparative alignments
  - Testing function of predicted promoters, enhancers using high-throughput reporter assays
- Linking cell-type specific expression to regulatory element structure
  - can we decipher the code of gene regulation?

Triaging candidate sequences for regulatory function

- Identify candidate regions from HSA19 comparative database
  - Further computations to identify additional elements, to map locations relative to known and predicted genes, to eliminate likely exons, and supplementary evidence e.g. Maps of transcription factor binding sites

- Design oligonucleotide primers, PCR and clone putative regions into commercially available reporter-construct vectors

- Transfect candidates into arrayed cell lines and assay for activity as promoters, enhancers

Test sequence Reporter gene (e.g. luciferase)

Sequence 1 (promoter) P E

Sequence 2 (enhancer) P E

Cell lines ------>

Zim1

Zim2

3 day-old mouse

Usp29

Znf264

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- Laurie Gordon

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- **Database design and sequence analysis:** Peg Folta, Astrid Terry, Carol Zhou, Qing Zhang, Sam Rash, Dan Rokhsar (JGI); Ed Uberbacher, Miriam Land (ORNL)

- **Mappers:** Anne Bergmann, Hummy Badri, Mari Christensen, Chi Ha, Sha Hammond, Matt Groza, Eddie Wehri, Michelle Vargas, Mark Wagner, Mark Shannon

http://www.jgi.doe.gov

For sequencing data (all is also in Genbank)

http://greengenes.llnl.gov/mouse/

For human map/ tiling path,
mouse BAC tiling path, restriction maps, accession numbers

On line soon:

•A catalog of known and predicted genes, with mouse conservation data
•comparative sequence alignments with parallel
sequence feature displays
•search tools for sequence match downloads